

# NxGen<sup>®</sup> T4 DNA Ligase (Low Concentration)



IMPORTANT!

**-20 °C Storage Required**

Immediately Upon Receipt

FOR RESEARCH USE ONLY. NOT FOR HUMAN OR DIAGNOSTIC USE.

## Table of Contents

Technical Support .....	2
Product Description .....	2
Product Specifications.....	3
Product Designations and Kit Components .....	3
Components & Storage Conditions.....	3
Reaction Set-Up .....	4
Related Lucigen Products .....	4
References .....	4
Notice of Limited Label License, Copyright, Patents, Warranties, Disclaimers and Trademarks .....	5

## Technical Support

Lucigen is dedicated to the success and satisfaction of our customers. Our products are tested to assure they perform as specified when used according to our recommendations. It is imperative that the reagents supplied by the user are of the highest quality. Please follow the instructions carefully and contact our technical service representatives if additional information is necessary. We encourage you to contact us with your comments regarding the performance of our products in your applications. Thank you.

### Lucigen Technical Support

Email: [techserv@lucigen.com](mailto:techserv@lucigen.com)

Phone: (888) 575-9695

Product Guarantee: Lucigen guarantees that this product will perform as specified for one year from the date of shipment.

## Product Description

T4 DNA Ligase catalyzes the formation of a phosphodiester bond between the terminal 5' phosphate and a 3' hydroxyl groups of duplex DNA or RNA. The enzyme efficiently joins blunt and cohesive ends and repairs single stranded nicks in duplex DNA, RNA or DNA/RNA hybrids (1).

**Storage buffer:** T4 DNA Ligase is supplied in 10 mM Tris-HCl, 50 mM KCl, 1 mM dithiothreitol, 0.1 mM EDTA, 0.1% Triton X-100, 50% glycerol, pH 7.5 @ 25 °C.

**10X T4 DNA Ligase Buffer** is composed of 500 mM Tris-HCl, 100 mM MgCl<sub>2</sub>, 50 mM dithiothreitol, 10 mM ATP, pH 7.6 @ 25 °C.

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**Source of protein:** A recombinant *E. coli* strain carrying the cloned T4 DNA Ligase gene.

**Unit Definition:** One Weiss unit is defined as the amount of enzyme required to convert 1 nmol of <sup>32</sup>P-labeled inorganic pyrophosphate into Norit adsorbable material in 20 minutes at 37 °C, using specified reaction conditions(2).

Note: 1 Weiss Unit is approximately 67 cohesive end units.

## Product Specifications

TEST	SPECIFICATION
Purity (SDS-PAGE)	>99%
SS Exonuclease	6,000 U <0.1% released
DS Exonuclease	6,000 U <0.1% released
Endonuclease	6,000 U <0.1% converted
<i>E. coli</i> 16S rDNA Contamination	3,000 U <10 copies

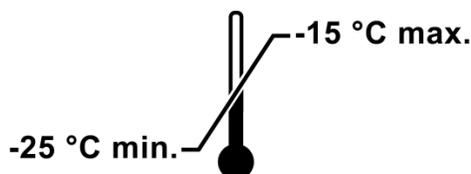
## Product Designations and Kit Components

Product	Ligase Concentration	Kit Size	Catalog Number	Reagent Description	Part Numbers	Volume
NxGen <sup>®</sup> T4 DNA Ligase (Low Concentration)	2 Units* / $\mu$ L	1,500 Units*	30241-1	T4 DNA Ligase (Low Concentration)	F83911-1	750 $\mu$ L
				10X T4 DNA Ligase Buffer	F88912-1	1.5 mL
		7,500 Units*	30241-2	T4 DNA Ligase (Low Concentration)	F83911-1	5 x 750 $\mu$ L
				10X T4 DNA Ligase Buffer	F88912-1	5 x 1.5 mL

\*Weiss Units

## Components & Storage Conditions

Store all Kits and Components at -20 °C



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## Reaction Set-Up

1. Add all of the components below to a clean reaction vessel.

Quantity	Component
2 $\mu$ L	10X T4 Ligation Buffer
X <sup>1</sup>	Vector
Y <sup>1</sup>	Insert
1 $\mu$ L	T4 DNA Ligase (2 U/ $\mu$ L)
To 20 $\mu$ L	Nuclease-free Water

<sup>1</sup>Recommended molar ratio of Vector to Insert is 1:3. Use 25-100 ng of vector and 75-300 ng of insert for each reaction.

2. Mix well by pipetting.
3. Incubate at 25 °C for 30 minutes.
4. Heat inactivate the reaction by incubating the ligation at 70 °C for 15 minutes.
5. Purify DNA using a PCR clean-up column and elute in ~50  $\mu$ L.

–OR–

Immediately dilute in TE or water (at least 1:10).

6. Transform 0.1-10 ng of the ligation product into a chemically competent or electrocompetent cell line that is compatible with the vector.

## Related Lucigen Products

NxGen<sup>®</sup> Phi29 DNA Polymerase (Catalog # 30021)

NxGen<sup>®</sup> M-MuLV Reverse Transcriptase (Catalog # 30222)

NxGen<sup>®</sup> T7 RNA Polymerase (Catalog # 30223)

NxGen<sup>®</sup> Lambda Exonuclease (Catalog # 30261)

NxGen<sup>®</sup> Exonuclease I (Catalog # 30262)

NxGen<sup>®</sup> Exonuclease III (Catalog # 30263)

## References

1. Engler, M. J., and Richardson, C. C. (1982) DNA ligases. In *The Enzymes*, Vol. XV (Ed. P. D. Boyer) Academic Press, New York, 3-29.
2. Weiss, B., Thompson, A., and Richardson, C. C. (1968) Enzymatic breakage and joining of deoxyribonucleic acid, VII. Properties of the enzyme-adenylate intermediate in the polynucleotide ligase reaction. *J. Biol. Chem.* 243, 4556-4563.

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